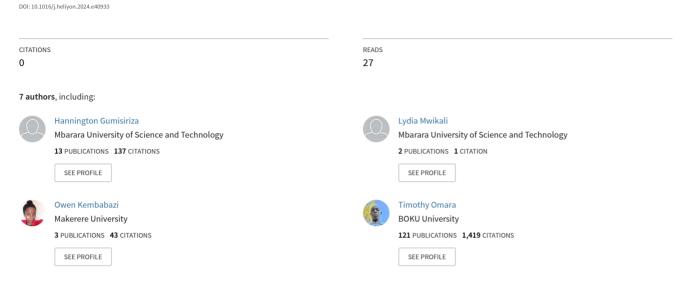
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Ethnopharmacology, phytochemistry, pharmacology and toxicity of the genus Gouania

Article in Heliyon · December 2024



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Review article

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Ethnopharmacology, phytochemistry, pharmacology and toxicity of the genus *Gouania*

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ARTICLE INFO

Keywords: Gouaniasides Gouanogenin Gouanic acids Gouania longipetala Acute toxicity

ABSTRACT

The genus Gouania (Rhamnaceae) comprises at least 50 recognized species distributed across tropical and subtropical regions. Gouania species have been ethnomedicinally used to treat a variety of ailments. Despite their widespread medicinal use, there is no comprehensive documentation that consolidates the ethnobotanical knowledge, phytochemicals, pharmacological properties, and toxicity of Gouania species. Herein, this review details the ethnopharmacology, phytochemical constituents, pharmacological properties and toxicity of Gouania species to provide perspectives for future research on this genus. Based on available literature, herbal preparations from Gouania species have been used to treat ailments related to the digestive, cardiovascular, respiratory, skin, musculoskeletal, reproductive, endocrine and urological systems. Extracts and isolated compounds from seven Gouania species (G. leptostachya, G. longipetala, G. lupulozdes, G. macrocarpa, G. longispicata, G. obtusifolia, and G. ulmifolia) have demonstrated promising anticancer, antimicrobial, antioxidant, and antiviral properties, supporting their ethnomedicinal uses. To date, 64 compounds (including 6 phenolic compounds, 24 flavonoids, and 34 terpenoids) have been isolated and characterized in the genus mainly as gouaniasides I-IX, gouanogenins, and gouanic acids. Most Gouania species remain unexplored for their potential bioactivities. The identification of more than 54 % as novel compounds from just seven Gouania species highlights the genus as a promising source for discovering new therapeutic agents to combat the growing challenge of multidrug-resistant pathogens. Conducting extensive phytochemical and pharmacological analyses across a broader array of Gouania species could unveil a more comprehensive profile of bioactive compounds, and pave way for innovative treatments against a diverse range of pathogens and diseases.

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https://doi.org/10.1016/j.heliyon.2024.e40933

Received 24 June 2024; Received in revised form 1 December 2024; Accepted 3 December 2024

Available online 5 December 2024

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1. Introduction

The Gouania Jacq. genus comprises of up to 70 distinct taxa in the buckthorn family (Rhamnaceae) [1]. The genus was established in 1763 by Jacquin based on two species from Haiti ("Domingensibus") namely: *G. tomentosa* and *G. glabra* [2,3]. The genus name commemorates Antoine Goüan (1733–1821), a pioneer in Linnaean taxonomy [4]. *Gouania* was classified as monophyletic within the tribe *Gouaniae* by Endlicher [5], a classification which was later reconfirmed by Richardson [6]. These plants are characterized as evergreen woody vines or climbing shrubs [7], with distinctive features that sets them apart from other genera in the family [8,9]. The plants feature alternate leaves, typically serrated with glandular teeth, and small persistent stipules, while the (bisexual) flowers are actinomorphic and arranged in spikes, racemes, or panicles. The flowers are characterized by bell-shaped hypanthium and

Table 1

Ethnobotanical uses of Gouania species based on published literature.

Species	Disease/conditions treated	Parts used	Preparation and administration	Country	References
Gouania longispicata Engl.	Foetal troubles, anorexia, oral thrush, stomachache, increasing strength in children, swelling in groin, boils, ulcers, malaria, allergy, febrile seizures, urinary retention, palpitations, heat rash, mastitis, syphilis, limb pains, syphilis, sweating, tooth decay, wounds, itching eyes, inflammations, colic pain, itching vagina, worms, hypertension, dizziness, asthma, body weakness, lactation insufficiency, typhoid, leprosy, leukoderma, hasten child birth, treat stomachache and malaria, lung and skin cancers, babesiosis and constipation in livestock	Sap, stem exudate, leaves, roots	Infusion taken orally, ointment, tea	Uganda, Tanzania, Rwanda, Cameroon, Ethiopia, Democratic Republic of Congo	[17,31–34, 57–64]
Gouania longipetala Hemsl.	Swelling, diabetes mellitus, gout, venomous stings, women infertility, cardiovascular diseases, snake bites, edema, venereal diseases, lumbago, malaria, pain, febrifuges, wounds, abdominal pain, cleaning of the born baby, body sweats, wounds, abdominal pain, genital stimulant, laxative, eye treatments, pain killer, heart diseases, lumbago, rickets, skin disorders, analgesic, sore eyes, conjunctivitis, iritis, ophthalmia, trachoma, gastro-intestinal problems, gynecological complaints	Stem, leaves, aerial parts, sap	Decoction/infusion taken orally, ointment, eye drops	Cameroon, South East Asia, Nigeria, Ghana	[16,18,20, 30,65–69]
Gouania lupuloides Urb.	Teeth cleaning, hypertension, malignant ulcers, pneumonia, depurative, diabetes, leishmaniasis	Root, stem, sap, bark	Decoction taken orally, brushing teeth, topical application	Jamaica, Cuba, Costa Rica, Mexico, Ecuador	[24,36,37, 40,43,70]
Gouania leptostachya DC.	Sores, numbness, stomachache, diarrhea, inflammations, postpartum herbal bath, food supplement, antispasmodic, fainting, leucorrhea	Leaves, stem, root	Poultice, cold infusion or decoction taken orally, bathing, smoke inhalation, potion	India, Indonesia, Thailand, Vietnam, Bhutan, Nepal	[13,41,44, 45,71–73]
Gouania macrocarpa DC.	Body pain	Root	Cold infusion mixed with rice gruel and taken orally	India	[13]
<i>Gouania javanica</i> Miq.	Oral thrush	Stem	Bubbles are blown to the thrush	Philippines	[46]
Gouania mauritiana Lam.	Obesity, diabetes, gastroenteritis and inflammations	Leaf, stem	Not reported	France	[21,47]
Gouania scandens (Gaertn.) R.B. Drumm. (Synonym: Gouania tiliifolia Lam.)	Emmenagogue, diuretic in dropsy	Not reported	Not reported	Mauritius	[48]
Gouania latifolia Reissek	Uterine inflammation and back pain	Bast, leaves	Not reported	Brazil	[49]
Gouania tiliifolia Lam. Gouania polygama (Jacq.) Urb.	Cough Hypertension, skin infections, stomachic, depurative and diuretic properties	Leaves Leaves	Crush decoction and fermentation of <i>G. polygama</i> with <i>Smilax</i> <i>domingensis</i> and <i>Pimenta dioica</i> with cane sugar	Madagascar Cuba, Mexico	[50] [51,52]
Gouania tiliaefolia Lam.	Liver disease, wounds, sores, ulcers, fever, headache	Leaves, whole plant	Leaf juice, poultice on sores	India, Bangladesh, Philippines	[53,74–77]
Gouania meyenii Steud.	Fever, headache, skin diseases	Not reported	Not reported	Hawaii	[15]

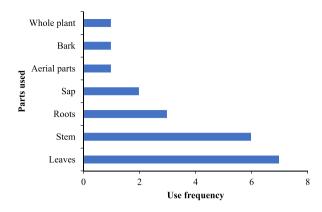


Fig. 1. Frequency of use of different parts of Gouania species for preparation of herbal remedies.

Table 2

Terpenoids reported from the genus Gouania.

Tetratriacontanoic acid (2)	Terpenoid	Species	Part used	References
Gouanic acid A (3)G. ulmifoliaAerial parts[14]Gouanic acid B (4)[14,27]Gouanic acid B (4)[27]Jozaciroside A (6)G. longipetala, G. leptostachya[1,27]Jozaciroside A (6)G. longipetala, G. leptostachya[27]Alphitolic acid (7)[25,27,8]Lupeol (8)[27]Betulinic acid (9)G. longipetala[27] β -sitosterol-3-O- β D-glucoside (10)G. lupulozdesStem[24]Gouanogenin A (11)G. lupulozdesStem[24]Gouanoside A (14)Guanoside A (14)Gouanoside B (15)Progouanogenin (16)3-O- β -D-glucopyranosyl gouanogenin A (17)G. longipetalaAerial parts[25]Gouaniside I (18)G. longipetalaAerial parts[25]Jujubosite I (19)G. longipetalaAerial parts[25]Gouaniside II (20)Gouaniside II (20)Ferrodovental (24)[26]Gouaniside II (20)Gouaniside II (21)[26][26]Gouaniside II (22)Gouaniside II (22)[26][26]Gouaniside II (23)G. leptostachya[82][26]Gouaniside IV (25)G. leptostachya[26][26]Gouaniside IV (28)[26][26][26]Gouaniside IV (28)[26][26][26]Gouaniside IX (29)Jazericoside (24)[26][26]Gouaniside IX (29)Jazericoside II (23)[26]Gouaniside IX (29)Jazericoside II (28)[26]Gouaniside IX (29)Jazericosid	Gouanic acid (1)	G. microcarpa	Leaves	[13]
Gouanic acid B (4) [14,27] 3-O,-D-p-glucopyranosyl Gouanogenin A (5) G. longipetala, G. leptostachya [27] Jozaciroside A (6) G. longipetala, G. leptostachya [1,27] Alphitolic acid (7) [27,82] Betulinic acid (9) G. longipetala [27] \$-sitosterol-3-O- \$-D-glucoside (10) [27] \$-gouanogenin B (12) [24] Bolin lactone (13) Gouanoside A (14) \$-gouanogenin (16) [25] \$-g-D-glucopyranosyl gouanogenin A (17) Gouaniside I1 (20) \$-gouaniside I1 (20) Gouaniside I1 (21) \$-gouaniside I1 (20) Gouaniside I1 (21) \$-gouaniside I1 (22) Gouaniside I1 (24) \$-gouaniside I1 (25) [26] \$-n-but/p-D-frucopyranoside (26) \$-leptostachya \$-gouaniside V1 (25) [26] \$-gouanisi	Tetratriacontanoic acid (2)			
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Lupeol (8) $[27,82]$ Betulin: acid (9) G. longipetala $[27]$ β -sitosterol-3-O- β D-glucoside (10) Gouanogenin A (11) G. lupulozdes Stem [24] Gouanogenin B (12) Ebelin lactone (13) Gouanoside A (14) Gouanoside A (14) Gouanoside B (15) Progouanogenin (16) $3-O_r\beta-D_rglucopyranosyl gouanogenin A (17)Gouaniside I (18) G. longipetala Aerial parts [25]Jujuboside I (19)Gouaniside I (19)Gouaniside II (20)Gouaniside II (20)Gouaniside II (21)Gouaniside II (22)Gouaniside IV (22)Gouaniside IV (23)Terminolic acid (24)Gouaniside VI (25)n-butyl-\beta-D-fructopyranoside (26) G. leptostachya [82]Gouaniside VI (27) G. leptostachya [82]Gouaniside IVI (28)Gouaniside IX (29)Jozeiroside C (30)Epigouanic acid A (31)Gouaniside IX (29)Gouanic acid A (32) G. leptostachya [82]Canothenic acid (33)$	Joazeiroside A (6)	G. longipetala, G. leptostachya		[1,27]
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Gouanioside A (32) G. leptostachya [82] Ceanothenic acid (33)	Joazeiroside C (30)			
Ceanothenic acid (33)	Epigouanic acid A (31)			
	Gouanioside A (32)	G. leptostachya		[82]
Daucosterol (34)	Ceanothenic acid (33)			
	Daucosterol (34)			

yellowish-green to whitish hood-shaped petals, with stamens often hooded by petals and arranged around a cup-shaped nectary disk. The ovary is inferior, with 3 locules each containing a single ovule while the fruit is a three-winged schizocarp, splitting into three indehiscent sections with shiny obovate seeds [3].

Gouania species are native to tropical zones of Africa (including Madagascar), Australia, Hawaii, Indian Ocean Islands, and Southern Asia [6,8,10,11], Mediterranean and Western Pacific [12]. A revision by Cahen [7] recognized five *Gouania* species in Philippines and Sundaland basing on morphological evidences (including a newly described species, *G. longipedunculata*). Another study reported that two species (*G. leptostachya* and *G. maderaspatana*) are endemic to India [13]. A taxonomic revision of the genus *Gouania* for Madagascar and the other Western Indian Ocean Islands recognized 17 distinct species, of which nine (9) were newly described [4]. Seventeen *Gouania* species were reported in Brazil, out of which only *Gouania* ulmifolia Hook. & Arn. grows in Rio Grande do Sul, but is also found in Southern Brazil, Uruguay and Argentina [14]. Since the last two decades, some *Gouania* species were cited to be critically endangered [8], which calls for establishment of conservation strategies and policies towards sustainability of such species. For instance, a single-island endemic *Gouania meyenii* discovered on Kauai in 1991 by Lorence & Flynn was last observed in 1994, and concerted efforts to relocate it were unsuccessful until the plants were rediscovered in 2020 by drone survey in Kalalau Valley, where conservation collections have been made [15].

Gouania species hold significant ecological and medicinal importance in their distributional ranges. They contribute to the biodiversity of ecosystems by providing habitat and food for animals, maintaining soil stability and preventing soil erosion. Members of this genus are renowned for their extensive bioactivities and thus use in ethnomedicine [16]. *Gouania* species feature prominently in traditional medicine where they are used to address cardiovascular, musculoskeletal, respiratory, skin, endocrine, urological, genital and digestive disorders [14,16–21]. *Gouania* species have appreciable antimicrobial [14,22], anti-inflammatory [23], antidiabetic, antilipidemic, antioxidant [18], and estrogenic effects [20] among other bioactivities.

The extensive ethnomedicinal applications of the genus *Gouania* is not surprising given its status as a promising source of novel constituents, with compounds like gouanogenins, gouaniasides, and gouanic acids, which are named after the genus [13,14,24–26]. Triterpenes are reported to be the most abundant phytochemicals in the *Gouania* genus, followed by flavonoids [25,27–29]. Studies on these compounds have exhibited diverse bioactivities including antioxidant, neuraminidase inhibition, anticancer, and antibacterial effects [25,27,28]. Despite their widespread medicinal use, there is no comprehensive documentation that consolidates the ethnobotanical knowledge, phytochemistry, pharmacology, and toxicity of the genus. Herein, this review collated literature on the ethnopharmacology, phytochemical constituents, pharmacological properties and toxicity of *Gouania* species to provide perspectives for future research on this genus.

2. Ethnomedicinal uses of Gouania species

Gouania species are used to treat up to 84 ailments, encompassing conditions affecting the digestive, cardiovascular, respiratory, skin, musculoskeletal, reproductive, endocrine and urological systems (Table 1). As an example, *G. longipetala* aerial parts (stems and leaves) are used in traditional medicine for the treatment of a variety ailments such as swelling, pain, gout, lumbago, venereal diseases, female infertility and cardiovascular diseases [27,30]. In Nigeria, *G. longipetala* has been used as genital stimulant and laxative [18]. The leaves of *G. longispicata* are used in Uganda to treat stomachache [17], opportunistic infections among HIV/AIDS patients [31] and over other 40 ailments including; allergy, and urinary retention [32]. In Ethiopia, *G. longispicata* is used to treat of oral thrush [33], leprosy and leukoderma [34].

The stems of *G. lupuloides* (Jamaican chewstick) have been used for teeth cleaning among African-Jamaicans for centuries [24,35]. Traditionally, a pencil-size piece of the bitter vine is chewed on one end to cause extensive foam and is then used to brush the teeth and massage the gums. As a result, *G. lupuloides* extracts have been in market as a mouthwash and dentifrice throughout Jamaica [24]. The Negroes grind stems of *G. lupuloides* in water and use it as antiseptic mouthwash or toothbrush [36]. The roots and stems of *G. lupuloides* are used to treat hypertension, while the sap is used to treat athlete's foot [37]. *Gouania lupuloides* is also used in traditional medicine in Cuba [38], Costa Rica [39] and Ecuador [40].

The leaves of *G. leptostachya* are used by the Lepcha (a tribe of Himalayan range living at the North-East corner of India) to make poultices for treating sores [13]. Leaves and stems of *G. leptostachya* are used by the Indonesians for treatment of stomachache and diarrhea [41]. The Mien of Northern Thailand use *G. leptostachya* as a postpartum herbal bath formula and food supplement, and also to treat convulsion in new born, numbness, fainting [42] and inflammations [23]. The rural folks of North Andaman, India burn the roots of *G. leptostachya* and then inhale the smoke to treat fever [43]. Paste made from the leaves of *G. leptostachya* is used to cure sores and inflammation in India [44,45].

The stem of *G. javanica* is used to treat fungal infections [46]. *Gouania mauritiana* is traditionally used to reduce cases of obesity and diabetes [21], gastroenteritis and inflammations [47]. *G. scandens* is used as an emmenagogue and diuretic in dropsy [48]. *G. latifolia* is used to treat uterine inflammation and back pain [49]. Leaf decoction of *G. tiliifolia* is orally taken in Madagascar for respiratory disorders [50]. *G. polygama* is used to make a traditional Cuban medicinal refreshment beverage produced by fermentation of a decoction of multispecies plants, chiefly with *Smilax domingensis, Pimenta dioica* and cane sugar [51]. *G. polygama* leaf is used to treat skin infections [52]. In India, *Gouania tiliaefolia* is used in Indian ethnomedicine for liver complaints [53], while the infusion from roots of *G. macrocarpa* is consumed to treat body pain [54]. Overall, leaves and stems are the most used parts of species of the *Gouania* genus for treatment of diseases (Fig. 1). The use of leaves may be related to their accessibility and abundance throughout the year, and their principal photosynthetic role and in the storage of therapeutic phytochemicals [55,56].

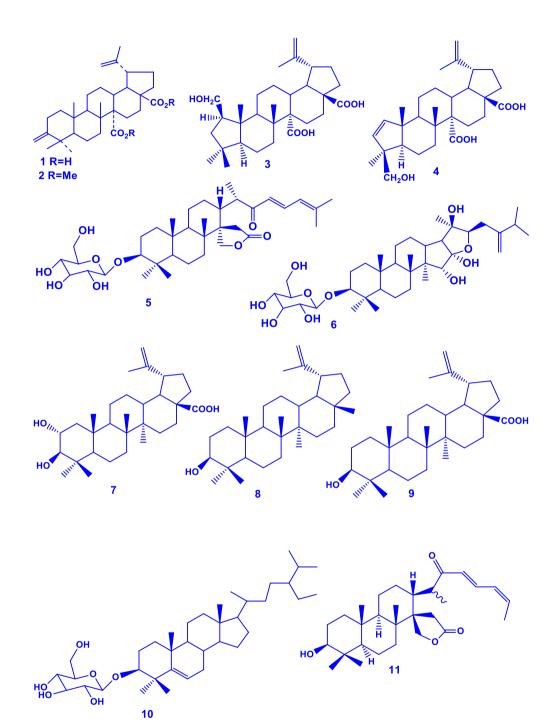


Fig. 2. Terpenoids characterized in Gouania species (1-34 correspond to the compounds in Table 2).

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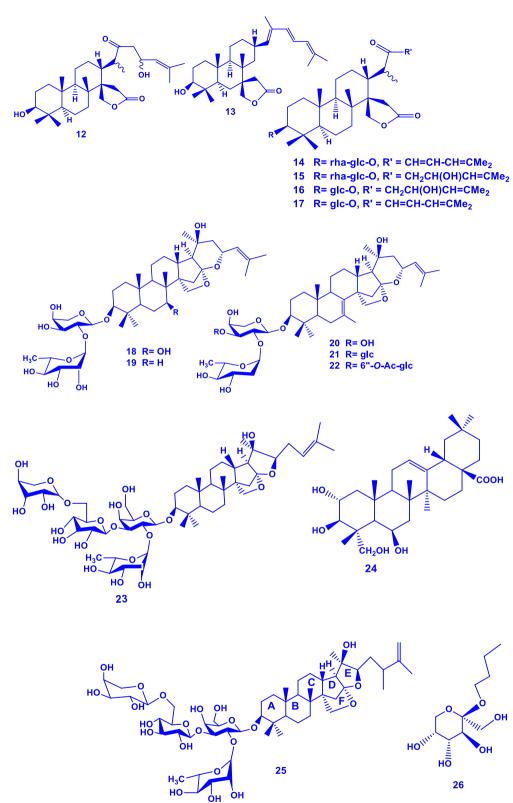


Fig. 2. (continued).

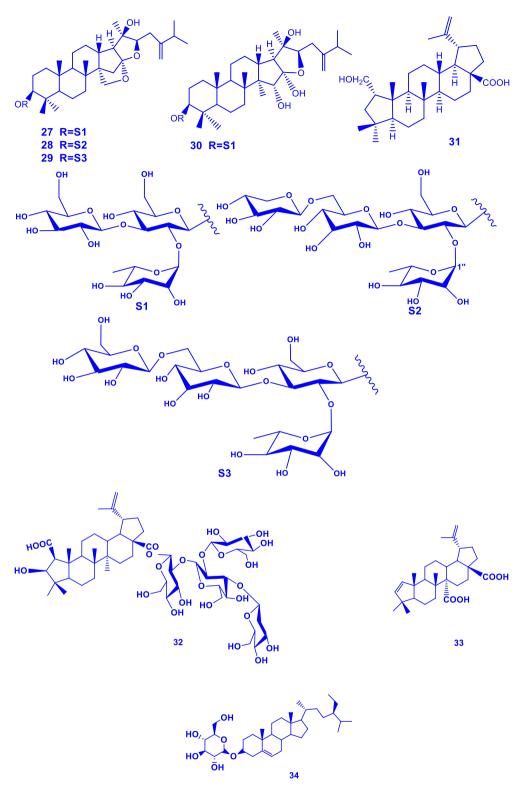


Fig. 2. (continued).

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Table 3

Phenolic compounds isolated from different parts of species in Gouania genus.

Phenolic compound	Species	Part	Reference
1-[(rel 2S,3R)-3,5,7-trihydroxy-3,4-dihydro-2H-chromen-2-yl]ethenone (35)	G. leptostachya	Stem	[85]
1-[(rel 2S,3S)-3,5,7-trihydroxy-3,4-dihydro-2H-chromen-2-yl]ethenone (36)	C lancinatala	A orial monto	[06]
Palmarumycin BG1/JC2 (37) de-O-methyllasiodiplodin (38)	G. longipetala	Aerial parts	[86]
Mangiferin (39)	G. obtusifolia	Leaves	[28]
4,6-dihydroxy-3-methylacetophenone-2-O-β-D-glucopyranoside (40)	G. longispicata	Leaves	[84]

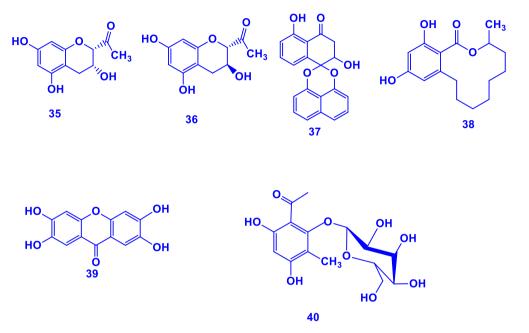


Fig. 3. Phenolic compounds characterized in Gouania species (35-40 correspond to the compounds in Table 3).

3. Phytochemistry of species in the Gouania genus

Several phytochemical screening reports have been made on members of the *Gouania* genus. For instance, *G. longipetala* stem ethanolic extract was reported to contain reducing sugars, triterpenoids, phenolics, phytosterols, saponins and flavonoids, with total phenolic content of 52.02 mg/g tannic acid equivalent [16]. Crude extracts of *G. longipetala* leaves contained alkaloids, phenols, saponins, flavonoids, steroids, tannins, terpenoids and cardiac glycosides with concentrations of 34.30, 23.19, 18.10, 16.49, 13.74, 9.72, 7.45 and 3.89 mg/100 g, respectively. Furthermore, GC-MS analysis of *G. longipetala* leaf extract revealed that it had 20 compounds. These included spartein (constituting up to 24.92 %), 2-tetradecanol, dodecanoic acid, 1-octadecene, luparine, 5,6-dehydrolupanine, propanoic acid-3-chloro-methyl ester, sapogenine, catechin, flavon-3-ol, anthocyanin, resveratrol, linoelaidic acid, aragyrine, methyl-9,12-heptadecadienoate, baptifoline, oleic acid, spartein, kaempferol and ethyl oleate [78].

Different solvent extracts of *G. longispicata* were qualitatively screened for phytochemicals and reportedly had cardiac glycosides, phenolics, steroids, flavonoids, saponins, and resins [22]. Another study reported that phenols, tannins, alkaloids and resins were present in *G. longispicata* methanolic leaf extract [79], while the whole plant extract contained polyphenols, flavonoids, triterpenes, sterols and saponins [80]. Similarly, *G. longispicata* leaf extracts had flavonoids, cardiac glycosides, saponins, steroids, resins and phenolics [22]. In a quantitative study, petroleum ether and methanolic extract of *G. tiliaefolia* had total phenolic content of 78.30 mg GAE/g and 70.37 mg GAE/g [77]. Phytochemical analysis of *G. longipetala* leaf extracts quantified vitamin C (2.807 mg/100 g), phenolics (0.375 mg GAE/g) and flavonoids (11.615 µg/g), while the corresponding quantities in the stem extract were 2.126 mg/100 g, 0.299 mg GAE/g and 10.012 µg/g [81]. Some phytochemical studies have been done to isolate and characterize bioactive compounds from *Gouania* species. These are discussed in the following subsections.

3.1. Terpenoids from the genus Gouania

Terpenoids have been reported to dominate in this genus, with up to 34 of them reported (**1-34**) (Table 2; Fig. 2). For example, *G. lupuloides* stem yielded two novel terpenoids (gouanosides A and B) which are 16,17-seco-dammaranoid saponins [24]. A new triterpenic acid (gouanic acid) was also identified in *G. macrocarpa* leaf extract [13]. In another report, gouanic acid A and B were characterized in *G. ulmifolia* aerial part extracts [14]. Six new triterpenoid saponins (named gouaniaside I-VI) were characterized from *G. longipetala* aerial parts [25]. Later, four novel such saponins (named gouaniasides VII–IX), along with joazeiroside C were identified in *G. leptostachya* aerial parts [26]. A unique and novel ceanothane-type saponin was similarly characterized in *G. leptostachya* [82]. More recently, an alkyl glycoside (*n*-butyl-β-D-fructopyranoside) was characterized from *G. leptostachya* [82].

3.2. Phenolic compounds from genus Gouania

A total of five phenolic compounds (**35–40**) have been reported in *Gouania* genus (Table 3; Fig. 3). Mangiferin (**39**), a well-known glucosylxanthone (xanthonoid) is the most important phenolic compound in this genus that was isolated from *G. obtusifolia* leaves [28]. This natural phenolic xanthonoid is known to possess excellent bioactivities such as antibacterial and anticancer activities [83]. A phenolic glycoside (named 4,6-dihydroxy-3-methylacetophenone-2-O- β -D-glucopyranoside (**40**)) was recently characterized from *G. longispicata* leaves [84].

3.3. Flavonoids from Gouania species

To date, it is known that *Gouania* species contain phenolic compounds, either as phenolics or flavonoids [28,29], signifying their prevalence in this genus. A total of 24 flavonoids (**41–64**) have been reported in this genus (Table 4; Fig. 4). These include rutin (quercetin-3-O-rutinoside), kaempferol, quercetin and their derivatives. Such flavonoids play essential roles in plant defense mechanisms, contributing to protection against ultraviolet visible radiation, pathogens, and herbivores [87].

Verily, alkaloids have been phytochemically identified among the secondary metabolites in *G. javanica* [46], *G. longipetala* [88], and *G. longispicata* [79]. However, no alkaloid has been isolated as a pure compound in any species of the *Gouania* genus. Therefore, future research could probe into the exploration and isolation of alkaloids within this genus. Despite the extensive traditional use of *Gouania* species reported in literature, characterization of bioactive compounds has been perfected in extracts from only 7 species (*G. longipetala*, *G. leptostachya*, *G. obtusifolia*, *G. macrocarpa*, *G. ulmifolia*, *G. longispicata* and *G. lupulozdes*). Specifically, *G. longipetala* has received the most attention in herbal medicine due to its widespread distribution compared to other *Gouania* species which grow in specific regions. Remarkably, more than 54 % of the yet characterized compounds from the *Gouania* genus are novel, suggesting its potential as a rich reservoir for discovering new drug leads to combat the growing array of pathogens. Therefore, extensive phytochemical investigations across a broader range of *Gouania* species beyond the seven investigated taxa could yield more compounds, which could act as scaffolds for development of potential drug candidates.

Table 4

Flavonoids isolated from the Gouania genus.

Flavonoids	Species	Part	Reference
Quercetin 3-O-α-L-rhamnoside (41)	G. obtusifolia	Leaves	[28]
Kaempferol 3-O-α-L-rhamnoside (42)			
Epicatechin (43)			
Gallocatechin (44)			
Engeletin (45)			
Kaempferol-3-O-(6-O-E-coumaroyl)- β -D-galactopyranosyl-(1 \rightarrow 2)- α -L-rhamnopyranoside (46)	G. longipetala	Aerial parts	[29]
Kaempferol-3-O-(6-O-E-feruloyl)- β -D-galactopyranosyl-(1 \rightarrow 2)- α -L-rhamnopyranoside (47)			
Kaempferol- 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-galactopyranoside (48)			
Kaempferol-3-O- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside (49)			
Kaempferol-3-O-α-L-rhamnopyranoside (50)			
Kaempferol-3- <i>O</i> - β -D-xylopyranosyl-(1 \rightarrow 2)- α -L-rhamnopyranoside (51)			
Kaempferol-3-O- β -D-galactopyr- anosyl- $(1 \rightarrow 2)$ - α -L-rhamnopyranoside (52)			
Quercetin-3-O- β -D-galactopyranosyl-(1 \rightarrow 2)- α -L-rhamnopyranoside (53)			
Quercetin-3-O- α - L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-galactopyranoside (54)			
Quercetin-3-O- β -D-xylopyranosyl-(1 \rightarrow 2) α - L- rhamnopyranoside (55)	G. leptostachya	Leaves	[1]
Quercetin-3-O-6-E-p-coumaroyl- β -D-glucopyranosyl- $(1 \rightarrow 2)$ - α -L-rhamnopyranoside (56)			
Quercitrin (57)	G. leptostachya	Aerial parts	[82]
Isoquercitrin (58)			
Kaempferol-3-O-(6-O-E-caffeoyl)- β -D-galactopyranosyl-(1 \rightarrow 2)- α -L-rhamnopyranoside (59)			
Catechin (60)			
Kaempferol-3-O-α-rhamnopyranoside (61)	G. longispicata	Leaves	[84]
Rutin (62)			
Kaempferol (63)			
Quercetin (64)			

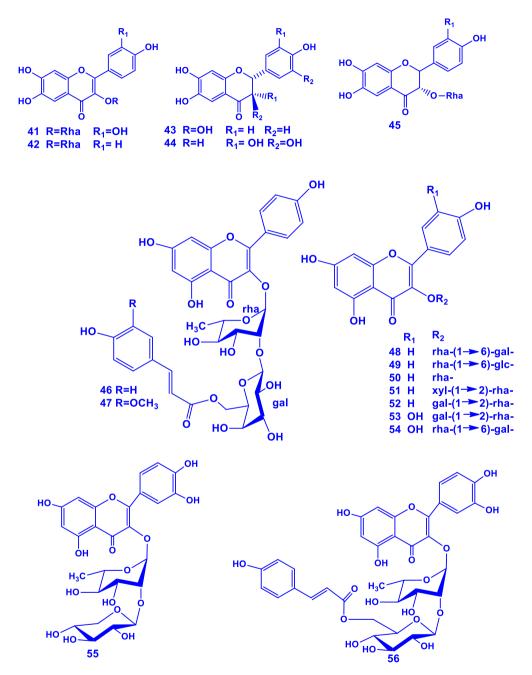
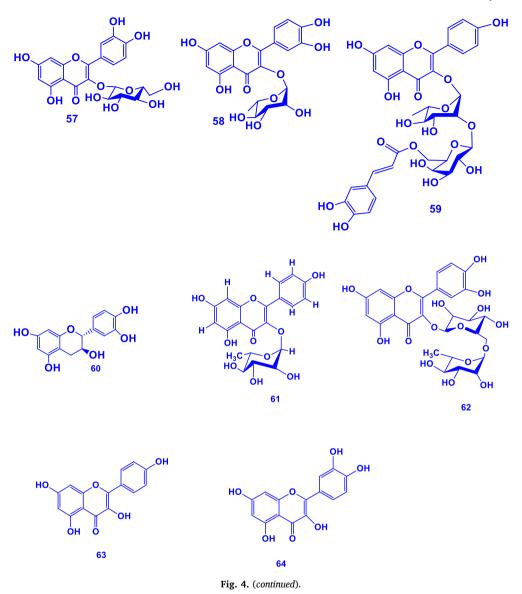


Fig. 4. Structure of flavonoids isolated and characterized in the Gouania taxon. The numbers 41-64 refer to the molecules mentioned in Table 4.



4. Bioactivity of extracts from Gouania species

4.1. Antioxidant activity

Ethanolic extract of *G. longipetala* stem had IC₅₀ of 0.004 mg/mL [16]. The DPPH assay of methanolic leaf extract of *G. longipetala* showed that it exhibited antioxidant activity (16.6 %–65.5 %) [88]. In another study, *G. longipetala* extract produced a dose-dependent and significant increase in superoxide dismutase and decrease in malondiadehyde [18]. *G. longipetala* fresh leaf extract lowered malondialdehyde concentration significantly [78]. Petroleum ether extract of *G. tiliaefolia* had free radical scavenging activity with IC₅₀ of 2.88 µg/mL, while the methanolic extract had IC₅₀ of 4.79 µg/mL [77].

4.2. Antimicrobial properties

Leaf extracts of *G. javanica* showed antifungal activity against *Aspergillus niger*, *Candida albicans* and *Tricophyton mentagrophytes* with mean zone of inhibition of 21.16 mm, 12.08 mm and 8.55 mm, respectively [46]. Ethanolic extract of *G. longipetala* stems

exhibited antibacterial effect against Bacillus subtilis, B. thurigiensis, Staphylococcus aureus, Enterococcus faecalis, Proteus vulgaris, Pseudomonas aeruginosa, Salmonella typhi and Escherichia coli, with the lowest MIC of $125 \mu g/mL$ against B. subtilis [16].

Recently, *G. longispicata* leaf extract inhibited growth of *S. aureus, Streptococcus pneumoniae, E. coli, P. aeruginosa, Candida albicans* and *Aspergillus flavus*, with *Streptococcus pneumoniae* being the most susceptible microorganism with MIC of 1.95 mg/mL and minimum bactericidal concentration of 15.625 mg/mL [22]. A similar study determined the antibacterial activity of nine Cameroonian medicinal plants against drug-resistant *E. coli, Enterobacter aerogenes, Klebsiella pneumoniae, Providencia stuartii, P. aeruginosa* and *S. aureus*, and the findings revealed that *G. longispicata* extract gave the broadest spectrum of action notably against 86.4 % of the bacterial strains tested [80]. Tannins, phenols, alkaloids and resins of *G. longispicata* were considered to be responsible for the potential activity against *S. aureus* and *Streptococcus pneumonia* and *P. aeruginosa* [79]. In another study, *G. lupuloides* was effective against *P. falciparum* K1 strain [39] and its stem extracts gave less than 50 % inhibition of *Mycobacterium tuberculosis* at 50 µg/mL [89].

4.3. Antidiabetic activity

Leaf extracts of *G. longipetala* significantly decreased the fasting blood sugar (FBS) levels of diabetic rats from 16.2 mM/L to 6.5 mM/L at 150 mg/kg within 24 h, and the FBS levels decreased by 62.0 %, 74.8 %, and 75.0 % on day 21 at 50 mg/kg, 100 mg/kg, and 150 mg/kg, respectively [88]. Using the oral glucose tolerance test, methanolic extract of *G. tiliaefolia* reduced blood glucose levels by 71.42 % and 75.39 % at doses 200 and 400 mg/kg-body weight, respectively compared with glibenclamide standard that reduced blood glucose levels by 66.17 % [77].

4.4. Anti-inflammatory activity

Ethanolic extract of *G. longipetala* stem bark had a maximal inhibitory effect of total edema by 93.78 % at 300 mg/kg [16]. A study assessing the anti-inflammatory effects of *G. leptostachya* extracts and fractions by inhibiting nitric oxide production in RAW 264.7 macrophages reported that the effects were concentration-dependent [26]. Another research group concluded that the ethanolic extract and fractions of *G. leptostachya* induced inflammatory effects by modulating nitric oxide synthesis in lipopolysaccharide-stimulated murine RAW264.7 macrophage cells, suggesting the potential therapeutic utility of *G. leptostachya* in inflammatory conditions [82].

4.5. Other bioactivities

There are other bioactivities of *Gouania* species. For example, the stem bark extract of *G. longipetala* has been shown to possess estrogenic properties [81]. Extracts of *G. tiliaefolia* showed insignificant thrombolytic and membrane stabilizing activities results compared to the respective standards [77]. Following castor oil-induced diarrhea, *G. tiliaefolia* methanol extract gave better reduction of diarrhea by 71.43 % (at 400 mg/kg-body weight) compared to loperamide (64.29 %) [66]. The central- and peripheral-analgesic activity of *G. tiliaefolia* methanol extract was dose-dependent compared to the standard, diclofenac sodium [77]. A study determined the anti-osteoclastogenesis of *G. javanica* leaf extracts and the results unveiled its potential as an effective inhibitor of osteoclastogenesis [90]. These components of *G. longipetala* exhibited various bioactivities including; inhibition of uric acid, amino acid decarboxylase activity, urine acidifiers, oligosaccharide provider, arachidonic acid inhibitor, decrease endothelial leukocyte and platelet adhesion [91].

5. Bioactivity of isolated compounds

Although 64 compounds have been characterized from *Gouania* genus, only 26 of these have been assessed for their bioactivities, such as antioxidant, anticancer, neuraminidase inhibition, and antibacterial properties (Table 5). Despite the documented isolation of compounds from this genus, scientific evaluation of efficacy, toxicity, and characterization of bioactive compounds has primarily focused on *G. longipetala*. Nevertheless, plants with significant ethnomedical usage have yielded potent compounds, with some integrated into essential medicines [92], which suggests that the genus *Gouania* can be a great source of new drug substances.

Among the ten (10) compounds from *G. longipetala* tested for effects on promyelocytic leukemia HL60 and human erythromyeloblastoid leukemia K562 cell lines, only jujuboside I demonstrated moderate cytotoxicity [25]. Additionally, the antibacterial activity of the isolated compounds against *Staphylococcus aureus, Escherichia coli*, and *Enterococcus faecalis* revealed alphitolic acid as the sole compound with significant inhibitory activity against all the three bacteria, giving MIC values ranging from 32 to 128 mg/mL [25]. The first study of bioactive compounds from *G. longispicata* [84], found that compounds from its leaves methanolic extract (40, **61–64**) exhibited antibacterial activity against *S. pneumoniae* and *E. coli* with MIC from 16 to 125 µg/mL. The radical scavenging activities of compounds (40, **61–64**) from *G. longispicata* leaves had IC₅₀ between 18.6 µg/mL and 28.1 µg/mL. The IC₅₀ of **63** and **64** were not significantly different from that of ascorbic acid, which indicated their promising utilization as natural antioxidants.

Table 5

Bioactivities of compounds isolated from genus Gouania.

Compound	Bioactivity tested	Species	Part used	Treatment/method	IC ₅₀ (μM)	References
Gouanic acid B (4) 3-O-β-D-glucopyranosyl gouanogenin A (5)	Antioxidant	G. longipetala	Aerial parts	DPPH method	52.5 56.4	[27]
Joazeiroside A (6) Alphitolic acid (7)					69.5 68.2	
Lupeol (8)					63.2	
Quercetin 3- O - α -L-rhamnoside (40)	Neuraminidase	G. obtusifolia	Leaves	200 µg/mL	7.89	[28]
Kaemferol 3- O - α -L-rhamnoside (41)	inhibition			(Neu5Ac2en control	5.47	
Epicatechin (42)				$IC_{50} = 44.47$)	5.49	
Gallocatechin (43)					10.22	
Engeletin (44)					5.44	
Mangiferin (39)					0.82	50.13
4,6-dihydroxy-3-	Antioxidant	G. longispicata	Leaves	DPPH method	26.3	[84]
methylacetophenone-2-O-β-D-	Antibacterial			Broth dilution	$MIC = 32 \mu g/mL$ and 64 $\mu g/mL$	
glucopyranoside (40)				method	mL	
Kaempferol-3- O -(6- O -E-feruloyl)- β -D-galactopyranosyl-(1 \rightarrow 2)- α -L-rhamnopyranoside (46)	Antioxidant	G. longipetala	Aerial parts	100 mg/mL; Ascorbic acid control ($IC_{50} = 60$)	47.4	[29]
Kaempferol- $3\text{-}O\text{-}\alpha\text{-}L\text{-}$ rhamnopyranosyl- $(1\rightarrow 6)\text{-}\beta\text{-}D\text{-}$ galactopyranoside (47)	Antioxidant				40.4	
Quercetin-3-O- α -L- rhamnopyranosyl-(1 \rightarrow 6)- β -D- galactopyranoside (53)					41.5	
Quercetin-3- O - β -D- galactopyranosyl- $(1 \rightarrow 2)$ - α -L- rhamnopyranoside (52)	Antioxidant	G. longipetala			13.8	
Jujuboside I (19)	Anticancer			HL60 cells	13.5	
Alphitolic acid (7)	Antibacterial activity			S. aureus, E. faecalis and E. coli (Gentamicin control MIC = 0.5)	MIC = 32, 64 and 128 mg/ mL, respectively	
Gouaniaside VII (27) Gouaniaside VIII (28) Gouaniaside IX (29)	Anti- inflammatory	G. leptostachya		NO inhibition by RAW 264.7 macrophages	The level of NO produced decreased in a concentration-dependent	[26]
Joazeiroside C (30)		0.1		NO.1111.1	manner	5003
Gouanioside A (32)	Anti- inflammatory	G. leptostachya		NO inhibition by RAW 264.7	The level of NO produced decreased in a	[82]
				macrophages	concentration-dependent manner	
Kaempferol-3-O-	Antioxidant	G. longispicata	Leaves	DPPH method	20.0	[84]
α-rhamnopyranoside (61)	Antibacterial	G. Wigspiculu	Leaves	Broth dilution method	MIC = 16 μ g/mL and 32 μ g/mL	[0]]
Rutin (62)	Antioxidant Antibacterial			DPPH method Broth dilution	28.1 MIC = 16 μg/mL and 32 μg/	
				method	mL	
Kaempferol (63)	Antioxidant Antibacterial			DPPH method Broth dilution	19.8 MIC $= 32 \ \mu\text{g/mL}$ and 125	
				method	µg/mL	
Quercetin (64)	Antioxidant			DPPH method	18.6	
	Antibacterial			Broth dilution method	$MIC = 64~\mu g/mL$ and $32~\mu g/mL$	

Note: IC₅₀ values are in µM. MIC, Minimum inhibitory concentration.

6. Toxicity profile of Gouania species

Few toxicity studies have been reported for the genus *Gouania*. Acute toxicity tests of *G. longipetala* extract suggested no observable signs of toxicity or morbidity, with LD_{50} greater than 4000 mg/kg [88], while a sub-chronic study showed that the relative liver weight increased significantly on day 90 at 10 mg/kg [18]. Additionally, changes in hematological parameters were not significant for *G. longipetala* extract-treated and untreated rats, suggesting that the extract was safe [18]. However, long term therapy of *G. longipetala* (up to 90 days) suggested toxicity to the liver and kidney, particularly at high doses. Another report found that *G. longipetala* extract significantly reduced total cholesterol, triglycerides and very low-density lipoproteins, while high density lipoproteins and serum urea increased prominently on day 90 at 10 mg/kg. At all doses of *G. longipetala* extract tested, liver enzyme markers increased significantly [18]. Acute toxicity study of *G. longipetala* leaf extract revealed a $LD_{50} > 5000 \text{ mg/kg}$ bw, signifying safe oral use of the plant. Also, sub-acute administration did not significantly alter the hematological, hepatic and renal function indices [69]. The petroleum ether

extract of *G. tiliaefolia* gave brine shrimp lethality $LC_{50} = 2.59 \mu g/mL$, while methanolic extract had LC_{50} of 3.38 $\mu g/mL$ [77]. These findings imply that *Gouania* species are safe for traditional use, however, further studies need to be done to give scientific evidence of their safety.

7. Conclusions and future research directions

Gouania species are utilized traditionally for treatment of digestive, cardiovascular, respiratory, skin, musculoskeletal, reproductive, endocrine and urological systems ailments. Only seven Gouania species (*G. leptostachya, G. longipetala, G. macrocarpa, G. lupulozdes, G. ulmifolia, G. longispicata* and *G. obtusifolia*) have been investigated to date and found to exhibit anticancer, antimicrobial, antioxidant, and antiviral activities. In total, 64 compounds have been so far characterized from *Gouania* genus but studies targeting their toxicity are still limited. Future research on *Gouania* species should investigate the phytochemicals and bioactivities of the unstudied species such as *G. scandens, G. mauritiana, G. latifolia, G. polygama, G. tiliaefolia* and G. *meyenii* that have been indicated for use in herbal formularies. Given the sparse data on the toxicity profiles of *Gouania* species, there is a critical need for systematic toxicity studies to ensure their safety when used in herbal medicine. Additionally, conducting clinical studies to assess the efficacy and safety of *Gouania* extracts and the characterized bioactive molecules could provide valuable insights into their medicinal properties and support the discovery of novel bioactive molecules.

CRediT authorship contribution statement

Hannington Gumisiriza: Writing – review & editing, Writing – original draft, Funding acquisition, Conceptualization. Eunice Apio Olet: Writing – review & editing, Supervision. Lydia Mwikali: Writing – review & editing. Racheal Akatuhebwa: Writing – review & editing, Data curation. Owen Kembabazi: Writing – review & editing. Timothy Omara: Writing – review & editing, Formal analysis. Julius Bunny Lejju: Writing – review & editing, Conceptualization.

Data availability

This review generated no raw data.

Funding

This review was funded by the Faculty of Science, Mbarara University of Science and Technology (Grant number: 7088522).

Declaration of competing interest

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:Hannington Gumisiriza reports financial support was provided by Mbarara University of Science and Technology, Faculty of Science. Hannington Gumisiriza reports a relationship with Mbarara University of Science and Technology, Faculty of Science that includes: funding grants. All authors have no conflict. If there are other authors, they declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

- N.T.K. Thuy, D.T. Trang, M. Thi, N. Trang, N.K. Bay, B.H. Tai, Flavonol glycosides and dammarane saponin from *Gouania leptostachya*, Vietnam J. Chem. 57 (3) (2019) 277–280, https://doi.org/10.1002/vjch.201960026.
- [2] N.J. Jacquin, Selectarum Stirpium Americanarum Historia. Ex Officina Krausiana, 1763. Vindobonae [Wien].
- [3] A. Pool, Taxonomic revision of Gouania (rhamnaceae) for north America, Ann. Mo. Bot. Gard. 99 (3) (2014) 490–552, https://doi.org/10.3417/2013016.
- [4] S. Buerki, P.B. Phillipson, M.W. Callmander, A taxonomic revision of *Gouania* (rhamnaceae) in Madagascar and the other islands of the western Indian ocean (the comoro and mascarene islands, and the Seychelles), Ann. Mo. Bot. Gard. 98 (2) (2011) 157–195, https://doi.org/10.3417/2007075.
- [5] S. Endlicher, Rhamneae: Genera Plantarum Secundum Ordines Naturales Disposita, Beck, Wien, 1840, pp. 1094–1105.
- [6] J.E. Richardson, M.F. Fay, Q.C.B. Cronk, M.W. Mashe, A revision of the tribal classification of Rhamnaceae, Kew Bull. 55 (2000) 311–340, https://doi.org/ 10.2307/4115645, 2014.
- [7] D. Cahen, K.S. Stenn, T.M.A. Utteridge, A revision of the genus Gouania (Rhamnaceae) in the Philippines and Sundaland, Kew Bull. 75 (4) (2020) 1–24, https:// doi.org/10.1007/S12225-020-9867-5.
- [8] D. Medan, C. Schirarend, Rhamnaceae. Flowering plants, Dicots 376 (1789) (2004) 320-338.
- [9] M.S. Sousa-Baena, N.R. Sinha, J. Hernandes-Lopes, L.G. Lohmann, Convergent evolution and the diverse ontogenetic origins of tendrils in angiosperms, Front. Plant Sci. 9 (403) (2018), https://doi.org/10.3389/fpls.2018.00403.
- [10] H. John, Monograph of the Hawaiian Species of Gouania (Rhamnaceae), Hawaiian Plant Stud. 96818 (1969) 507-543.
- [11] N.A. Jud, M.A. Gandolfol, A. Iglesias, P. Wilf, Flowering after disaster: early Danian buckthorn (Rhamnaceae) flowers and leaves from Patagonia, PLoS One 12 (5) (2017) e0176164, https://doi.org/10.1371/journal.pone.0176164.
- [12] H. Lieth, M.J.A. Werger, Ecosystems of the World. CSIRO Division of Land Resources Management, 1989. Wembley, W.A. (Australia).
- [13] S.P. Nair, J.M. Rao, Gouanic acid from the leaves of *Gouania microcarpa*, Phytochem. 33 (3) (1993) 711–712.
- [14] S.R. Giacomelli, G. Maldaner, C. Stücker, C. Marasciulo, J. Schmidt, L. Wessjohann, I.I. Dalcol, A.F. Morel, Triterpenoids from Gouania ulmifolia, Planta Med. 73 (2007) 499–501, https://doi.org/10.1055/s-2007-967166.
- [15] B. Nyberg, K.R. Wood, S.M. Deans, S. Heintzman, A.D.A.M. Williams, Recent notable plant records and rediscoveries from Kaua'i, Hawaiian Islands, Bish. Mus. Occas. Pap. 148 (2023) 163–168, https://doi.org/10.1055/s-0032-1328326.

- [16] E. Ekuadzi, R.A. Dickson, T.C. Fleischer, Antibacterial, anti-inflammatory and antioxidant properties of Gouania longipetala hemsl, Int. J. Pharm. Sci. 3 (5) (2012) 1300–1305.
- [17] F.A. Hamill, S. Apio, N.K. Mubiru, M. Mosango, R. Bukenya-Ziraba, O.W. Maganyi, D.D. Soejarto, Traditional herbal drugs of southern Uganda, J. Ethnopharmacol. 70 (2000) 281–300, https://doi.org/10.1016/s0378-8741(00)00180-x.
- [18] M.I. Ezeja, A.O. Anaga, I.U. Asuzu, Acute and sub-chronic toxicity pro fi le of methanol leaf extract of *Gouania longipetala* in rats, J. Ethnopharmacol. 151 (3) (2014) 1155–1164, https://doi.org/10.1016/j.jep.2013.12.034.
- [19] S. Kokate, Wonderful Plants Combating Skin Diseases, 2014.
- [20] P. Désiré, M. Ngadena, Y. Sandrine, B. Danielle, K. Mireille, A.O. Bibi-farouck, D. Théophile, K. Pierre, *In vivo* estrogenic-like activities of *Gouania longipetala* Hemsl. (Rhamnaceae) bark extracts in a post-menopause-like model of ovariectomized Wistar rats, J. Ethnopharmacol. 168 (2015) 122–128, https://doi.org/ 10.1016/j.jep.2015.03.049.
- [21] M. Marimoutou, F.S. Sage, J. Smadja, C.L. d'Hellencourt, M. Gonthier, C.R. Silva, Antioxidant polyphenol-rich extracts from the medicinal plants Antirhea borbonica, Doratoxylon apetalum and Gouania mauritiana protect 3T3-L1 preadipocytes against H₂O₂, TNFα and LPS inflammatory mediators by regulating the expression of superoxide dismutase and NF-κB genes, J. Inflamm. 12 (2015) 10, https://doi.org/10.1186/s12950-015-0055-6.
- [22] H. Gumisiriza, G. Birungi, J.B. Lejju, E.A. Olet, O. Kembabazi, C.D. Sesaazi, Ethnobotany and antimicrobial activity of *Gouania longispicata* engl, J. Complement. Med. Res. 11 (1) (2020), https://doi.org/10.5455/jcmr.2020.11.01.10.
- [23] M.T.T. Dung, J. Lee, E. Kim, B.C. Yoo, V.T. Ha, Y. Kim, D.H. Yoon, S. Hong, K. Baek, N.Y. Sung, T.W. Kim, J. Kim, J.Y. Cho, Anti-inflammatory activities of Gouania leptostachya methanol extract and its constituent resveratrol, Phytother Res. 29 (3) (2014) 381–392, https://doi.org/10.1002/ptr.5262.
- [24] E.J. Kennelly, W.H. Lewis, R.E.K. Winter, S. Johnson, M. Elvin-Lewis, J. Gossling, Triterpenoid saponins from Gouania lupuloides, J. Nat. Prod. 56 (3) (1993) 402–410, https://doi.org/10.1021/np50093a013.
- [25] M.A.A. Gossan, P. Yao-Kouassi, A.A. Coffy, J. Josse, S.C. Gangloff, H. Morjani, L. Voutquenne-nazabadioko, Triterpene glycosides from the aerial parts of Gouania longipetala, Phytochemistry (Elsevier) 134 (2016) 71–77, https://doi.org/10.1016/j.phytochem.2016.11.004.
- [26] N.T. Hang, T.N.T. Bich, V. LeBa, O.T. Van, N.M. Khoi, T.H. Do, Characterisation of four new triterpenoid saponins with nitric oxide inhibitory activity from aerial parts of *Gouania leptostachya*, Nat. Prod. Res. 36 (23) (2022) 5999–6005, https://doi.org/10.1080/14786419.2022.2057971.
- [27] S. Désiré, N. Ernestine, T.B. Bruno, S. Sidjul, D.D. Ulrich, M. Lateef, B. Schneider, S. Ali, N. Barthélemy, A new dammarane type triterpene glucoside from the aerial parts of *Gouania longipetala* (Rhamnaceae), Nat. Prod. Res. 35 (19) (2019) 3192–3203, https://doi.org/10.1080/14786419.2019.1690483.
- [28] X. Li, T. Ohtsuki, S. Shindo, M. Sato, Mangiferin identified in a screening study guided by neuraminidase inhibitory activity, Planta Med. 73 (2007) 37–38, https://doi.org/10.1055/s-2007-981582.
- [29] M.A.A. Gossan, A. Yao-kouassi, A.A. Coffy, D. Harakat, L. Voutquenne-Nazabadioko, New acylated flavonol glycosides from the aerial parts of *Gouania longipetala*, Phytochem. Lett. 11 (2015) 306–310, https://doi.org/10.1016/j.phytol.2015.01.019.
- [30] M.D. Yemele, P.B. Telefo, L.L. Lienou, S.R. Tagne, C.S.P. Fodouop, C.S. Goka, M.C. Lemfack, F.P. Moundipa, Ethnobotanical survey of medicinal plants used for pregnant women' s health conditions in Menoua Division-West Cameroon, J. Ethnopharmacol. 160 (2015) 14–31, https://doi.org/10.1016/j.jep.2014.11.017.
- [31] G. Anywar, E. Kakudidi, R. Byamukama, J. Mukonzo, A. Schubert, H. Oryem-origa, Indigenous traditional knowledge of medicinal plants used by herbalists in treating opportunistic infections among people living with HIV/AIDS in Uganda, J. Ethnopharmacol. 246 (2020) 112205, https://doi.org/10.1016/j. jep.2019.112205.
- [32] H. Gumisiriza, G. Birungi, E.O. Olet, C.D. Sesaazi, Medicinal plant species used by local communities around queen elizabeth national park, maramagambo central forest reserve and Ihimbo central forest Reserve, South western Uganda, J. Ethnopharmacol. 239 (2019) 111926, https://doi.org/10.1016/j. jep.2019.111926.
- [33] M. Giday, Z. Asfaw, Z. Woldu, Ethnomedicinal study of plants used by Sheko ethnic group of Ethiopia, J. Ethnopharmacol. 132 (1) (2010) 75–85, https://doi. org/10.1016/j.jep.2010.07.046.
- [34] N. Tuasha, B. Petros, Z. Asfaw, Medicinal plants used by traditional healers to treat malignancies and other human ailments in Dalle District, Sidama Zone, Ethiopia, J. Ethnobiol. Ethnomed. 14 (2018) 1–21, https://doi.org/10.1186/s13002-018-0213-z.
- [35] S.A. Mitchell, The Jamaican root tonics: a botanical reference, Focus on CAM 16 (4) (2011) 271–280, https://doi.org/10.1111/j.2042-7166.2011.01124.x.
- [36] J. Soelberg, O. Davis, A.K. Jäger, Historical versus contemporary medicinal plant uses in the US Virgin Islands, J. Ethnopharmacol. 192 (2016) 74–89, https:// doi.org/10.1016/j.jep.2016.07.005.
- [37] R.L. Kuby, Folk Medicine on St. Croix; an Ethnobotanical Study, The University of Kansas, United States, 1979.
- [38] E. Caamal-Fuentes, L.W. Torres-Tapia, P. Simá-Polanco, S.R. Peraza-Sánchez, R. Moo-Puc, Screening of plants used in Mayan traditional medicine to treat cancer-like symptoms, J. Ethnopharmacol. 135 (2011) 719–724, https://doi.org/10.1016/j.jep.2011.04.004.
- [39] J. Ferrier, A. Saleem, A.C. Ramirez, R. Liu, E. Chen, T. Pesek, V. Cal, M. Balick, J.T. Arnason, Traditional medicines used by Q'eqchi' Maya to treat diabetic symptoms and their antiglycation potential, J. Ethnopharmacol. 224 (2018) 504–511, https://doi.org/10.1016/j.jep.2018.06.031.
- [40] M.S. Gachet, J.S. Lecaro, M. Kaiser, R. Brun, H. Navarret, R.A. Munoz, R. Bauer, W. Schuhly, Assessment of anti-protozoal activity of plants traditionally used in Ecuador in the treatment of leishmaniasis, J. Ethnopharmacol. 128 (2010) 184–197, https://doi.org/10.1016/j.jep.2010.01.007.
- [41] P.W. Grosvenor, P.K. Gothard, N.C. McWilliam, A. Supriono, D.O. Gray, Medicinal plants from Riau province, Sumatra, Indonesia. Part 1: uses,
- J. Ethnopharmacol. 45 (1995) 75–95, https://doi.org/10.1016/0378-8741(94)01209-1.
- [42] K. Panyaphu, P. Sirisa-ard, P.N. Ubol, S. Nathakarnkitkul, S. Chansakaow, O.T. Van, Phytochemical, antioxidant and antibacterial activities of medicinal plants used in Northern Thailand as postpartum herbal bath recipes by the Mien (Yao) community, J. Phytopharmacol. 2 (1) (2012) 92–105.
- [43] P.R.C. Prasad, C.S. Reddy, S.H. Raza, C.B.S. Dutt, Folklore medicinal plants of north Andaman Islands, India, Fitoterapia 79 (6) (2008) 458–464, https://doi. org/10.1016/j.fitote.2008.03.007.
- [44] B.K. Pradhan, H.K. Badola, Ethnomedicinal plant use by Lepcha tribe of Dzongu valley, bordering Khangchendzonga Biosphere Reserve, in north Sikkim, India, J. Ethnobiol. Ethnomed. 4 (2008) 1–18, https://doi.org/10.1186/1746-4269-4-22.
- [45] Y. Uprety, R.C. Poudel, J. Gurung, N. Chettri, R.P. Chaudhary, Traditional use and management of NTFPs in Kangchenjunga Landscape: implications for conservation and livelihoods, J. Ethnobiol. Ethnomed. 12 (2016) 1–59, https://doi.org/10.1186/s13002-016-0089-8.
- [46] A.B. Segismundo, P.E. Florendo, A.R.P. Pablico, In vitro antifungal activity and phytochemical screening of Gouania javanica Miq, Leaves, IJESTM 17 (1) (2008), https://doi.org/10.69566/ijestm.v17i1.210.
- [47] C. Poullain, E. Girard-Valenciennes, J. Smadja, Plants from reunion island: evaluation of their free radical scavenging and antioxidant activities, J. Ethnopharmacol. 95 (1) (2004) 19–26, https://doi.org/10.1016/j.jep.2004.05.023.
- [48] S. Suroowan, K.B. Pynee, M.F. Mahomoodally, A comprehensive review of ethnopharmacologically important medicinal plant species from Mauritius, South Afr. J. Bot. 122 (2019) 189–213, https://doi.org/10.1016/j.sajb.2019.03.024.
- [49] U.P. Albuquerque, P.M. de Medeiros, A.L.S. Almeida, J.M. Monteiro, E.M. Neto, J.G. de Melo, J.P. dos Santos, Medicinal plants of the caatinga (semi-arid) vegetation of NE Brazil: a quantitative approach, J. Ethnopharmacol. 114 (2007) 325–354, https://doi.org/10.1016/j.jep.2007.08.017.
- [50] N.H. Rakotoarivelo, F. Rakotoarivony, A.V. Ramarosandratana, V.H. Jeannoda, A.R. Kuhlman, A. Randrianasolo, R.W. Bussmann, Medicinal plants used to treat the most frequent diseases encountered in Ambalabe rural community, Eastern Madagascar, J. Ethnobiol. Ethnomed. 11 (2015) 1–16, https://doi.org/10.1186/ s13002-015-0050-2.
- [51] M. Al Mamun, T.V. Gonzalez, A. Islam, T. Sato, S. Sato, T.K. Ito, M. Setou, Analysis of potential anti-aging beverage Pru, a traditional Cuban refreshment, by desorption electrospray ionization-mass spectrometry and FTICR tandem mass spectrometry, JFDA 27 (4) (2019) 833–840.
- [52] P.M. Bork, M.L. Schmitz, M. Kuhnt, C. Escher, M. Heinrich, Sesquiterpene lactone containing Mexican Indian medicinal plants and pure sesquiterpene lactones as potent inhibitors of transcription factor NF-κB, FEBS Lett. 402 (1) (1997) 85–90, https://doi.org/10.1016/s0014-5793(96)01502-5.
- [53] S.K. Jain, Ethnobotany and research on medicinal plants in India, National botanical research Institute, Novartis Found. Symp. 185 (1994) 153-168.
- [54] S. Binu, Medicinal plants used for treating body pain by the tribals in Pathanamthitta district, Kerala, India, IJTK 10 (2011) 547-549.

- [55] R.M.A. Machado, I. Alves-Pereira, R.M.A. Ferreira, Plant growth, phytochemical accumulation and antioxidant activity of substrate-grown spinach, Heliyon 4 (9) (2018) e00751, https://doi.org/10.1016/j.heliyon.2018.e00751.
- [56] B. Mwamatope, D. Tembo, E. Kampira, C. Maliwichi-Nyirenda, V. Ndolo, Seasonal variation of phytochemicals in four selected medicinal plants, Pharmacogn. Res. 13 (4) (2021), https://doi.org/10.5530/pres.13.4.14.
- [57] T. Tanno, Plant utilization of the Mbuti pygmies: with special reference to their material culture and use of wild vegetable foods, Afr. Stud. Monogr. 1 (1981) 1–53.
- [58] R. Irakiza, M. Vedaste, B. Elias, B. Nyirambangutse, N.J. Serge, N. Marc, Assessment of traditional ecological knowledge and beliefs in the utilisation of important plant species: the case of Buhanga sacred forest, Rwanda, Koedoe 58 (1) (2016) 1–11.
- [59] A. Kudamba, J.N. Kasolo, G.S. Bbosa, A. Lugaajju, H. Wabinga, N. Niyonzima, M. Ocan, A.M. Damani, H.M. Kafeero, J.E. Ssenku, S.O. Alemu, M. Lubowa, A. Walusansa, H. Muwonge, Medicinal plants used in the management of cancers by residents in the Elgon Sub-Region, Uganda, BMC Complement Med Ther. 23 (1) (2023) 450, https://doi.org/10.1186/s12906-023-04273-5.
- [60] F.M.O. Olsthoorn, Determinants of Spatial Patterns of Human Activity in Bwindi Impenetrable National Park (Doctoral Dissertation), Imperial College London, UK, 2017.
- [61] J.R. Tabuti, C.B. Kukunda, D. Kaweesi, O.M. Kasilo, Herbal medicine use in the districts of Nakapiripirit, Pallisa, Kanungu, and Mukono in Uganda, J. Ethnobiol. Ethnomed. 8 (2012) 1–35, https://doi.org/10.1186/1746-4269-8-35.
- [62] K. Chifundera, Livestock diseases and the traditional medicine in the Bushi area, Kivu province, Democratic Republic of Congo, Afr. Stud. Monogr. 19 (1) (1998) 13–33.
- [63] M. Kyoshabire, E. Katuura, A.B. Cunningham, R. Hoeft, Medicinal plants and herbalist preferences around Bwindi Impenetrable National Park, J. Med. Plants Res. 11 (2017) 161–170, https://doi.org/10.5897/JMPR2016.6317.
- [64] N.P. Mollel, J.N. Otieno, D.K. Sitoni, Medicinal plants traded in Arusha city, Tanzania, J Med Plants Stud. 10 (1) (2022) 175-182.
- [65] H.M. Burkill, The Useful Plants of West Tropical Africa, Royal Botanical Gardens, Kew, UK, 1985.
- [66] D. Abbiw, Useful Plants of Ghana, Intermediate Technical Publications and Royal Botanic Gardens, UK, 1990, p. 207.
- [67] P.J. Houghton, I.M. Osibogun, Flowering plants used against snakebite, J. Ethnopharmacol. 39 (1993) 1–29, https://doi.org/10.1016/0378-8741(93)90047-9.
- [68] D.A. Focho, W.T. Ndam, B.A. Fonge, Medicinal plants of Aguambu-Bamumbu in the Lebialem highlands, southwest province of Cameroon, Asian J. Pediatr. Pract. 3 (1) (2009) 1–13.
- [69] D. Njamen, M.A. Mvondo, S. Djiogue, G.J.M.K. Wanda, C.B.M. Nde, G. Vollmer, Phytotherapy and women's reproductive health: the Cameroonian perspective, Planta Med. 79 (7) (2013) 600–611.
- [70] J.H. Cano, G. Volpato, Herbal mixtures in the traditional medicine of Eastern Cuba, J. Ethnopharmacol. 90 (2004) 293–316, https://doi.org/10.1016/j. jep.2003.10.012.
- [71] K. Panyaphu, O.T. Van, P. Sirisa-Ard, P. Srisa-Nga, S. ChansaKaow, S. Nathakarnkitkul, Medicinal plants of the Mien (Yao) in Northern Thailand and their potential value in the primary healthcare of postpartum women, J. Ethnopharmacol. 135 (2) (2011) 226–237, https://doi.org/10.1016/j.jep.2011.03.050.
- [72] K. Srithi, C. Trisonthi, P. Wangpakapattanawong, H. Balslev, Medicinal plants used in Hmong women's healthcare in northern Thailand, J. Ethnopharmacol. 139 (1) (2012) 119–135, https://doi.org/10.1016/j.jep.2011.10.028.
- [73] A. Patra, A.K. Mukherjee, Assessment of snakebite burdens, clinical features of envenomation, and strategies to improve snakebite management in Vietnam, Acta Trop. 216 (2021) 105833, https://doi.org/10.1016/j.actatropica.2021.105833.
- [74] A.K. Khatun, M.A.A. Khan, M.A. Rahman, M.S. Akter, A.H. Abid Hasan, R.J. Ripa, M. Moniruzzaman, M.J. Mahal, M. Rahmatullah, Ethnomedicinal usage of plants and animals by folk medicinal practitioners of three villages in Chuadanga and Jhenaidah districts, Bangladesh, Am-Eur. J. Sustain. Agr. 7 (2013) 319–339.
- [75] M.A. Motaleb, M.M. Abdullah-Al-Mamun, M.K. Hossain, M. Alam, M. Sultana, Herbal healing: an old practice for healthy living among Khumi, Marma and Tripura communities of Thanchi Upazila, Bangladesh, Eur. J. Med. Plants 5 (1) (2015) 23–52, https://doi.org/10.9734/EJMP/2015/10244.
- [76] A.K.M.G. Sarwar, Medicinal plant genetic resources of Bangladesh-genera represented by single species and their conservation needs, J Med Plants Stud 3 (2) (2015) 65–74.
- [77] T. Ahmed, R. Khan, N. Tabassum, F. Aktar, M.K. Islam, Evaluation of bioactivities of *Gouania tiliaefolia* Lam., an Indigenous traditional medicinal plant of Bangladesh, Bangladesh Pharm. J. 22 (2) (2019) 200–207, https://doi.org/10.3329/bpj.v22i2.42305.
- [78] K.C. Nwachukwu, G.S. Aloh, S.I. Egbachukwu, S.N. Ijioma, L.A. Alli, K.C. Ugwuanyi, B.U. Onwuchekwa, M.P. Okoh, Phytochemical composition, GC-MS analysis and toxicological profiling of *Gouania longipetala* leaf extract in rats, Afr. J. Biotechnol. 22 (9) (2023) 192–201, 0.5897/AJB2023.17594.
- [79] H. Migabo, C. Izere, T. Habyarimana, U.K.N. Josiane, E. Nsabayezu, F.N. Niyonzima, Phytochemistry analysis and anti-bacterial properties of Rwandese medicinal plants, NeuroQuantology 20 (17) (2022) 965. https://orcid.org/0000-0002-8228-5171.
- [80] O.M.F. Demgne, C.F. Tchinda, A.T. Mbaveng, V.P. Beng, V. Kuete, Antibacterial and antibiotic-potentiating activities of nine Cameroonian medicinal plants against multidrug-resistant bacteria expressing active efflux pumps, Invest Med Chem Pharmacol 5 (1) (2022) 58, https://doi.org/10.31183/imcp.2022.00058.
- [81] D. Soh, A.T. Choumessi, F.P.T. Manfo, M.N.N. Ngum, M. Mbouh, E.A. Nantia, Antioxidant content and activity of stem and leaf extracts of *Gouania longipetala* Hemsl, Rhamnaceae), AYU 43 (2) (2022) 6064, https://doi.org/10.4103/ayu.AYU_241_18.
- [82] N.T. Hang, N.T.B. Thu, L.B. Vinh, N.V. Phong, O.T. Van, K.Y. Lee, A new ceanothane-type triterpenoid saponin isolated from *Gouania leptostachya* DC. var. tonkinensis Pit. and its underlying anti-inflammatory effects, JMB. 33 (7) (2023) 941, https://doi.org/10.4014/jmb.2301.01040.
- [83] A.H. Rahmani, A. Almatroudi, K.S. Allemailem, H.O.A. Alharbi, W.M. Alwanian, B.A. Alhunayhani, A.A. Khan, Role of mangiferin in management of cancers through modulation of signal transduction pathways, Biomed 11 (12) (2023) 3205, https://doi.org/10.3390/biomedicines11123205.
- [84] H. Gumisiriza, E.A. Olet, L. Mwikali, R. Akatuhebwa, T. Omara, J.B. Lejju, D.C. Sesaazi, Antibacterial and antioxidant activities of flavonoids, phenolic and flavonoid glycosides from Gouania longispicata leaves, Microbiol. Res. 15 (4) (2024) 2085–2101, https://doi.org/10.3390/microbiolres15040140.
- [85] C. Yao, S.J. Zhang, Z.Z. Bai, T. Zhou, L.J. Xuan, Two new benzopyran derivatives from Gouania leptostachya, Chin. Chem. Lett. 22 (2) (2011) 175–177, https:// doi.org/10.1016/j.cclet.2010.09.028.
- [86] E. Ekuadzi, R.A. Dickson, T.C. Fleischer, I.K. Amponsah, D. Pistorius, L. Oberer, Chemical constituents from *Gouania longipetala* and Glyphaea brevis, Nat. Prod. Res. 28 (15) (2014) 1210–1213, https://doi.org/10.1080/14786419.2014.921685.
- [87] L.B.D.S. Nascimento, M. Tattini, Beyond photoprotection: the multifarious roles of flavonoids in plant terrestrialization, Int. J. Mol. Sci. 23 (9) (2022) 5284, https://doi.org/10.3390/ijms23095284.
- [88] M.I. Ezeja, A.O. Anaga, I.U. Asuzu, Antidiabetic, antilipidemic, and antioxidant activities of Gouania longipetala methanol leaf extract in alloxan-induced diabetic rats, Pharm. Biol. 53 (4) (2015) 605–614, https://doi.org/10.3109/13880209.2014.935864.
- [89] J.G. Graham, S.L. Pendland, J.L. Prause, L.H. Danzinger, J.S. Vigo, F. Cabieses, N.R. Farnsworth, Antimycobacterial evaluation of Peruvian plants, Phytomedicine 10 (6) (2003) 528–535, https://doi.org/10.1078/094471103322331502.
- [90] L.T. Huong, D.T. Trang, N.H. Dang, Screening for Vietnamese plant extracts with potential benefit for anti-osteoclastogenesis, VJST 59 (4) (2021) 507–517,
- https://doi.org/10.15625/2525-2518/59/4/15825.
 [91] A.J. Madubuike, K.K. Igwe, I.E. Otuokere, F.J. Amaku, Bioactivity evaluation study of phytochemicals in *Gouania longipetala* ethanol leaf extract using GC-MS analysis, IJSTRE 1 (5) (2016) 63–71.
- [92] C. Veeresham, Natural products derived from plants as a source of drugs, \J. Adv. Pharm. Technol Res 3 (4) (2012) 200–202, https://doi.org/10.4103/2231-4040.104709.